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Characterization and Evaluation of Copper and Nickel Biosorption on Acidic Algae *Sargassum Filipendula*

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The marine algae *Sargassum filipendula* was collected from São Paulo seashore (Brazil) and submitted to treatment with acid. The biosorption mechanisms of Cu^{2+} and Ni^{2+} ions onto acidic algae *Sargassum filipendula* were examined using various analytical techniques: Fourier-transform infrared (FT-IR) spectroscopy, scanning electron microscopy (SEM), energy dispersive X-ray spectroscopy (EDX) and potentiometric titration (pH_{ZPC}). The effect of acidic treatment on algae by hydrochloric acid (pH 2.0, 3.0, 4.0 and 5.0) was evaluated for Cu^{2+} and Ni^{2+} adsorption. Alginate was extracted from raw algae and the two types of acids present in the biomass (β-D-mannuronic (M) and α-L-guluronic (G) acid) were characterized by ^{13}C NMR. The M/G ratio was found to be 0.50. According to the pH_{ZPC} analysis, at a pH higher than 5.5 the acidified algae surface presents a negative charge. The FT-IR analyses showed that the main chemical groups involved in the biosorption were carboxylic, alcoholic, sulfonate and amino groups.

Keywords: *^{13}C NMR, FT-IR, MEV/EDX, alginate, acid treatment*

1. Introduction

Environmental biotechnology studies have been conducted using alternative materials in order to eliminate heavy metal ions from industrial effluents. Biosorption is a cost effective process that uses inactive biosorbents for decontamination of pollutants, such as heavy metal ions[1]. This process consists in using materials of biological origin, more specifically living or dead microorganisms, to accumulate solute on the surface of the sorbent[2]. Many different kinds of biomass can effectively uptake heavy metals (algae, moss, fungi, bacteria, chitosan and zeolite)[2-11], but recent research has shown that marine algae are very effective[8,12] for metal binding.

Seaweed cells have a large superficial area with sites that are able to provide fast and reversible bonding with cations[13]. *Sargassum* sp. is a widespread and common kind of marine brown algae that has been used for metal recovery, due to the high content of polysaccharides in the cell wall, which are responsible for the high sorption capacity[14].

This material presents a high organic leaching when used for metal waste treatment. The consequences of the organic leaching can be many and would hinder its industries applications. This can lead to a secondary pollution and also to a decrease in the adsorption performance in water and wastewater treatment[14]. Therefore, it is important to modify the material before its application in biosorption processes. This modification can be done by using acid, base, calcium, or aldehyde[14-16].

Chen and Yang[14], studied the surface modification by formaldehyde or glutaraldehydes can result in a significant reduction of organic leaching in cationic biosorption and the biosorption capacity is enhanced due to the modifications and the biosorption kinetics is not affected.

There are several important functional groups in marine algae, such as carboxyl, sulfonic, and amino, which interact with metal ions through different mechanisms, including ion exchange, surface complex formation, micro precipitation, chelation, and coordination[17].

Raize et al.[18], using XPS analysis showed that cadmium cations bind to chemical groups possessing oxygen and carbon (carboxyl groups in the alginic acid), nitrogen (amino/amido groups in the peptidoglycans and proteins), and sulfur (sulfonate, thiol in the sulfate polysaccharides and amino acids).

The carboxylic groups are the most abundant acidic functional group and the adsorption capacity of algae is directly related to the presence of these sites in alginate. Alginic acid occurs in all brown algae and corresponds to 45% of the dry weight, corresponding to 2.25 mmol of carboxyl groups/g of biomass[19]. It is located in the intracellular matrix as a gel containing Na^+, Ca^{2+}, Mg^{2+}, Sr^{2+}, and Ba^{2+} ions[20]. Its main function is believed to be skeletal, giving both strength and flexibility to the algal tissue.

Mannuronic and guluronic acid residues have carboxylic and hydroxyl functional groups that take part in metal binding to alginate[18,21-23] studied the *Sargassum* metal (Cd^{2+}, Ni^{2+} and Pb^{2+}) binding capacities, before and after partial extraction of sulfate polysaccharides (fucoidan) and alginic acid. After extraction, the metal binding capacities decreased by 25, 55, and 75% for lead, cadmium, and nickel, respectively.

These carboxylic groups can be found in a linear (1→4)-linked-copolymer of β-D-mannuronic (M) and α-L-guluronic (G) acids. These two acids can be arranged in homopolymeric (MM) and (GG) or heteropolymeric (MG) sequences (blocks)[24].

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Both the polymer conformation (M and G in the alginate matrix) and the respective proportion of these groups may change with the a) kind of algae; b) the part of the plant where the polysaccharide is extracted; and c) the age or stage in algae growth.

Information about the sequential structure of alginates was obtained early by Haug et al.[24], and Haug and Smidsrod[25]. They concluded that alginate is a true block copolymer composed of homopolymeric regions of M and G. They found marked differences among the M/G ratios of alginates from different brown algae.

Variation in the affinity of some divalent metals to alginates with different M/G ratios was demonstrated early on by Haug et al.[24], and Haug and Smidsrod[25]. He showed that the affinity of alginates for divalent cations such as Pb^{2+}, Cu^{2+}, Cd^{2+}, Zn^{2+}, Ca^{2+}, etc. increased with the guluronic acid content. The selectivity coefficients for the ion-exchange reaction between sodium and divalent metals were determined for two alginates[25] and confirmed the higher affinity of guluronic acid rich alginates for divalent metals.

Brown algae are an abundant source for alginate extraction and this biopolymer is commercially available[23]. Commercial alginates are produced mainly from *Laminaria hyperborea, Macrocystis pyrifera, Laminaria digitata, Ascophyllum nodosum, Laminaria japonica, Eclonia maxima, Lessonia nigrescens, Durvillea Antarctica,* and *Sargassum* sp.[26].

The quality of the alginate extracted from *Sargassum* species found along the Brazilian coast is inferior and cannot be compared with the quality of the alginate extracted from species of *Ascophyllum, Laminaria* and *Macrocustis*[27]. However, the authors point to the possibility that the alginate extracted from *Sargassum* species found in the Brazilian southwest region may be enough to supply part of the domestic market demand.

A complete chemical characterization of brown algae substrate is necessary to emphasize the advantages of biosorption in relation to the conventional technique of ion-exchange using resins and other materials[1]. This work presents the characterization of *Sargassum filipendula* algae by solid state ^{13}C NMR nuclear magnetic resonance spectroscopy, in order to determine the M/G ratio in the alginate extracted from algae matrix. This M/G ratio is important to choose a biosorbent for heavy metal treatment. This study also investigated the influence of acid treatment on algae for copper and nickel adsorption. Techniques such as FT-IR spectroscopy, scanning electron microscopy (SEM) with energy dispersive X-ray spectroscopy (EDX) and pH_{ZPC} analysis were used to characterize the biomass, in its raw form, after acidic treatment and metal Cu^{2+}, Ni^{2+} binding.

2. Methods

2.1. Sargassum filipendula marine algae

The *Sargassum filipendula* marine algae was collected in São Paulo seashore (São Sebastião, Brazil) by the Biology Institute of São Paulo University (CebiMar). The samples were washed and rinsed in distilled water and stored at –20 °C. Algae samples were dried at 60 °C overnight and stored in a dry cabinet. The biomass was ground and sieved and fractions measuring from 0.71 to 1.0 mm were collected for further experiments.

2.2. Extraction of alginate from marine algae

The method described by Percival and McDowell[28] was used for the alginate extraction. The dry algae was soaked in formaldehyde for 24 hours at 60 °C, washed with water and dropped into HCl solution (0.2 mol.L^{-1}) for 24 hours. After this time the samples were washed again in distilled water before extraction in 2% solution of sodium carbonate. In the presence of an excess of Na_2CO_3, the alginic acid is converted to a sodium alginate causing polymer dissolution[29]. Alginate extraction was carried out at 60 °C by soaking for 3 hours. The samples were centrifuged and the supernatants were labeled as crude extracts. The alginate samples were then obtained from the crude extract by precipitation with ethanol. The precipitate was washed twice with acetone and freeze-dried.

2.2.1. Solid state MAS ^{13}C NMR nuclear magnetic resonance spectroscopy

After the alginate extraction procedure the presence of the two types of acids (i.e. guluronic acid (G) and mannuronic acid (M)), were determined using Solid state ^{13}C NMR Nuclear Magnetic Resonance spectroscopy, recorded with a Bruker Avance DRX 500 spectrometer (Bruker, Karlsruhe, Germany).

Solid state NMR of alginate samples (MAS ^{13}C NMR) is a technique that brings the advantage that sodium alginate powder can be examined directly, without need of partial hydrolysis. Normally this partial hydrolysis is used to decrease viscosity of high-molecular-weight alginate solutions prior to recording their proton NMR spectra. This step that can induce precipitation of a portion of alginate may significantly distort the experimental results[30].

2.3. pH_{ZPC} analyses

The procedure described by Davranche et al.,[31] was used to determine the pH of zero point charge of *Sargassum filipendula* algae. It was determined according to the surface complexation model, as described by Stumm[32]. Two algae solutions were prepared by immersing 5 g of algae samples in 100 mL of CH_3COONH_4 (0.1 mol.L^{-1}) as the supporting electrolyte. Both solutions were titrated with CH_3COOH (0.3 mol.L^{-1}) and NH_4OH (0.25 mol.L^{-1}).

The titrations were carried out over a wide range of pH. The total surface charge, Q, was calculated as a function of pH by Equation 1.

$$Q = \frac{C_A - C_B + [OH^-] - [H^+]}{W_s} \quad (1)$$

C_A and C_B are the acid and base concentrations (mol.L^{-1}), respectively, $[H^+]$ and $[OH^-]$ are the equilibrium concentrations of these ions (mol.L^{-1}), and W_s is the solid concentration (g.L^{-1}).

The pH_{ZPC} of the solid concentration can be estimated by drawing a curve of the total surface charge as function of pH. The pH_{ZPC} is the pH value where the curve crosses the x-axis (Q = 0).

2.4. Cu^{2+} and Ni^{2+} speciation

Diagrams of Cu^{2+} and Ni^{2+} species distribution as pH functions were simulated using HYDRA (Hydrochemical Equilibrium-Constant Database) software[33]. These diagrams were made for metal at equilibrium concentration (4 mmol.L^{-1}) that corresponded to the maximum copper and nickel adsorption.

2.5. Acid treatment of Sargassum filipendula marine algae and copper and nickel bioadsorption

A significantly high amount of organic leaching has been observed in the treatment and recovery of heavy metals by raw biosorbents, with a consequent elevation of pH. The pH elevation can cause a) metal precipitation; b) metal complexation by soluble ligands released from the algae, which may reduce the metal binding ability of biosorbents.

In this study, biomass particles were pre-treated with hydrochloric acid (pH 2.0, 3.0, 4.0 and 5.0) and pH was continuously adjusted, maintaining it close to the required values, until stabilization was achieved.

Algae samples, 0.5 g of raw or acid samples, were loaded with 100 mL of copper and nickel ions (250 mg.L^{-1}), pH 4.5. The

solubility of metal ions, under selected experimental conditions, was systematically checked. The suspensions were kept in a rotary shaker at 175 rpm for 6 hours. The heavy metal ions concentration was measured by inductive coupled plasma-emission spectroscopy (ICP-ES).

2.6. Fourier transforms infrared spectroscopy (FT-IR)

FT-IR spectroscopy was used to confirm the presence of the functional groups in samples of *Sargassum filipendula* and to observe the chemical modification after heavy metal adsorption in raw and acidified algae. Infrared spectra were recorded in the 4000-600 cm^{-1} region using a Thermo Nicolet instrument, model IR-200. The ATR (attenuated total reflection) device allows getting information about the surface.

2.7. SEM/EDX

The surface morphology of algae was observed using scanning electron microscope (SEM). After drying, the samples (0.71 to 1.0 mm) were covered with a thin layer of gold (10 nm) using a sputter coater (SCD 0050 – Baltec, Liechenstein) and observed using the JEOL JXA-840^{A} scanning electron microscope (20 kV) under vacuum of 1.33×10^{-6} mBar (Jeol, Japan). To determine the chemical composition, Energy dispersive X-ray spectroscopy was performed on algae after metal adsorption and acidic treatment. The samples were prepared as for SEM analyses. The objective of this analysis was qualitative, not quantitative.

3. Results and Discussion

3.1. Alginate extraction and characterization (Solid state MAS ^{13}C NMR spectroscopy)

To characterize the relation M/G in alginate matrix, this material was initially extracted from algae. The complex spectral pattern can be represented as a sum of individual symmetric signals corresponding to the C(2) – C(5) atoms of both uronic residues[34], enough to estimate the M/G ratio with high accuracy[30], assuming that the stronger, downfield peak is due to the main constituent, the α-L-guluronic acid.

The alginate composition can be determined by using the relation of total area: G-4+G-2+G-3+G-5 (guluronic) for M-4+M-2+M-3+M-5 (mannuronic) (Table 1). The M/G ratio was found to be 0.50.

This relation is a very important parameter to choose an alga for application in heavy metal treatments. It varies according to the extraction methodology and the location from which the algae was collected[29], as well as according to seasonal and growth conditions[35]. The comparison of the M/G ratio from other *Sargassum* species is given in Table 2.

Table 2 indicates different M/G rations for *S. fluitans*: 0.19[29], 0.52[37] and 1.18[19]. For *S. filipendula:* 0.19[38] and 0.50 (This study). These differences can be attributed due to changes in geographical locations and season of algae collect.

It has been reported[40] that most *Sargassum* alginates have M:G ratios ranging from 0.8 to 1.5. This will be discussed in more detail below and, as indicated in preceding sections of this review, low M:G ratios (i.e. < 1.0) are indicative of higher G content and are, therefore, deemed highly advantageous for the implementation of the biosorption process. This reflects the established selectivity for divalent cations of the guluronic block sections, in accordance with the ''egg-box'' model[1].

Many studies show high affinity of the *Sargassum* species, from different locations, in the removal of copper and nickel heavy metals: for copper values of 0.93 and 0.89 $mmol.g^{-1}$ for *S. vulgare* and *S. filipendula*, respectively[41], of 0.99[42] and 1.062[43] for *Sargassum* sp., had been found. For nickel, values of 0.75, 0.41 and 0.09 $mmol.g^{-1}$ for *S. fluitans*, *S. natans*, and *S. vulgare* respectively[44] and 0.61 for *Sargassum* sp.[16]; and 0.32 for *S. wightii*[45]. There are few published papers that compare brown algae regarding the presence of mannuronic and guluronic (M/G ration) acids with the removal capacity of heavy metals, which makes comparison difficult.

Considering that the main responsible for metal biosorption from brown seaweed species is biopolymer alginate, the determination of relation M/G in this biopolymer can be a basic factor in the election of the material for application as biosorbent in the heavy metals biosorption process.

3.2. pH_{ZPC} analysis

Carboxylic, amino and sulfate groups are the main binding sites in brown algae for metal adsorption. These groups can be ionized when pH varies. At low pH values, surface sites are protonated and the surface becomes positively charged, whilst the ionizable groups lose their protons and the surface becomes negatively charged at high pH values. Figure 1 depicts the charge variation in function of pH for in nature and acidified (pH 5.0) algae. For the in nature algae the pH_{ZPC} values were within the range of pH 6.0 and 7.0, and for the acidified algae (pH 5.0) the pH_{ZPC} values were within the range of pH 5.5 and 6.0. This means that at pH higher than 7.0 and 6.0, algae surface bears negative charges, for in nature and acidified algae, respectively. This difference is decurrently of the composite leaching gifts in the biomass (with consequent increasing of pH) before the acid treatment. This explains why the binding of many metals increases with increasing pH.

The pH dependence of metal biosorption can be explained by the fact that there is a competition among H^+ with heavy metal ions through a combination of mechanisms: ion exchange and the formation of surface metal complexes[14]. At low pH, this competition is strong, hence the metal uptake is lower. When pH is increased, the competitive effect becomes less important and more heavy metal ions are removed.

However, the adsorption mechanism is determined not only by the functional groups on the sorbents but also the characteristics of metal solutes. Both metal speciation in the solution and the functional groups on the biosorbents are relevant to metal binding mechanisms[1].

3.3. Cu^{2+} and Ni^{2+} speciation

Figure 2a and b show the distribution of copper and nickel species as a pH function for metal at equilibrium concentration of 4 $mmol.L^{-1}$ for Cu^{2+} and Ni^{2}, respectively.

It was observed that Cu^{2+} precipitate occurs above pH 5.0 with formation of $CuO_{(cr)}$. In the case of Ni^{2+}, precipitation occurs above pH > 6 with formation of $Ni(OH)_{2(c)}$. In order to perform the adsorption study of these metals, it is necessary to keep the pH below 5.0 for copper ions and about 6.0 for nickel ions.

3.4. Pre-treatment of Sargassum filipendula marine algae and biosorption of Ni^{2+} and Cu^{2+}

Marine algae contain a high amount of organic substances, such as carbohydrates, protein, lipids, and pigments. Some of these compounds may be leached from the biomass and can be released to the aqueous phase during the biosorption operation[46,47]. It is common to observe that after biosorption water changes to a yellowish or green color.

The effect of acidic treatment on algae by hydrochloric acid (pH 2.0, 3.0, 4.0 and 5.0) was evaluated for copper and nickel

Table 1. Peak intensity obtained by analysis of MAS ^{13}C resonances attributed to residues of α-L-guluronic (G) and β-D-mannuronic acid (M) in sodium alginate of *Sargassum filipendula.*

	M peaks				G peaks				M/G ratio
	M-5	M-4	M-3	M-2	G-5	G-4	G-3	G-2	
Peak (ppm)	81.3	83.0	79.4	75.8	68.8	83.3	71.0	66.0	0.50
Intensity	2.9	3.3	3.0	5.1	12.5	3.4	7.7	5.1	

Table 2. M/G ratio of alginate materials extracted from different *Sargassum* species.

Sargassum species	Origin	M/G	Reference
S. vulgare	Brazil (Northern)	1.27	36
S. fluitans	Cuba	0.52	37
S. oligocystum	Australia	0.62	
S. multicum	England	0.31	38
S. thunbergii	Korea	0.53	
S. oligocystum	Australia	0.77	
S. polycystum	Unknown origin	0.21	
S. filipendula	Unknown origin	0.19	
S. dentifolium	Egypt	0.52	39
S. asperifolium		0.69	
S. latifolium		0.82	
S. fluitans	Florida	1.18	19
S. fluitans	Cuba	0.19	29
S. siliquosum		0.72	
S. muticum	England	0.31	
S. filipendula	Brazil (Southeast)	0.50	This study

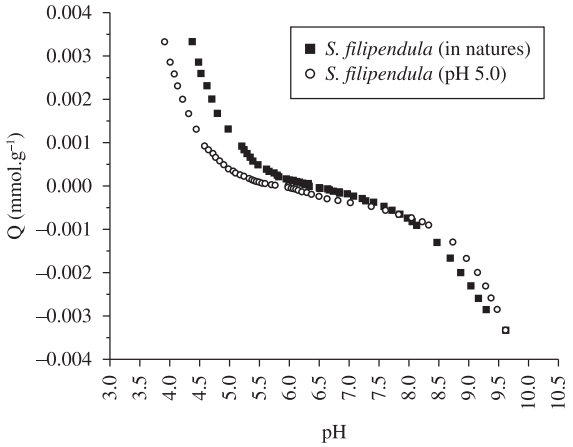


Figure 1. Total surface charge of in nature and acidified (pH 5.0) *Sargassum filipendula* algae as function of pH.

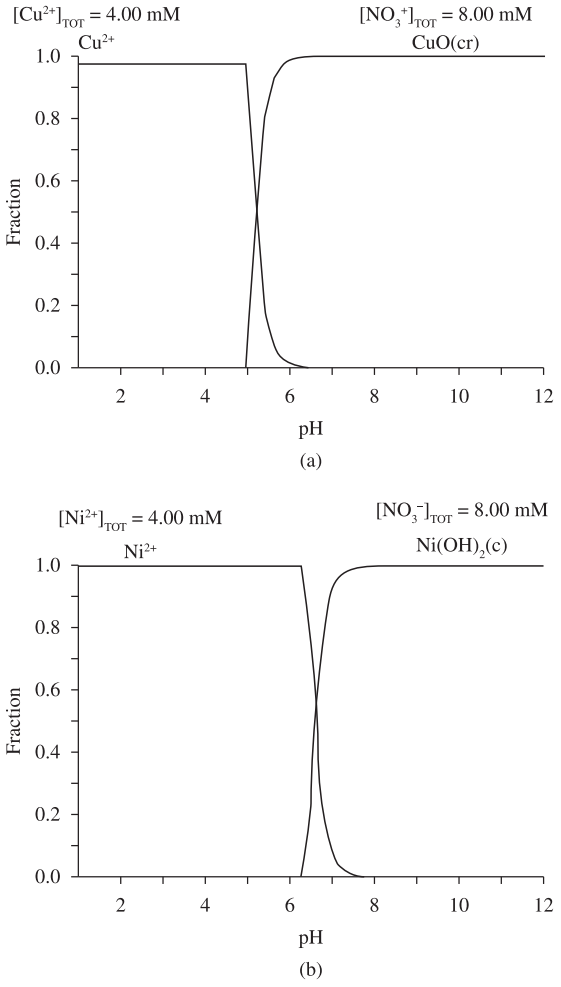


Figure 2. a) Cu^{2+}; and b) Ni^{2+} speciation in adsorption as a function pH.

adsorption (Figure 3a and b). A slight variation in copper and nickel removal was observed when compared to raw algae, indicating a decrease of pH from 5.0 to 4.0. For a pH variation from 3.0 to 2.0, an average decrease of 30% was observed for both copper and nickel.

The highest effect in adsorption properties at low pHs can be explained by the chemical modification in the active sites, resulting from the treatment with acid. A weight loss of 27 and 26% was observed at pH 2.0 and 3.0 and 23 and 22% at pH 5.0 and 4.0, respectively.

It is interesting to observe that the decrease in sorption efficiency is not correlated to weight loss. Almost the same weight loss occurred at pH 5 and 2, while the sorption capacity remained unchanged at pH 5, whereas it decreased by 40-50% at pH 2 and 3. This probably means that this weight loss is not responsible foe the decrease in sorption capacity. This may be explained by the pH effect on the interaction of metal ions with binding groups (both on the sorbent and or the compounds leached).

At lower pH, the concentration of H^+ ions is higher, leading to a lower sorption of metals ions. These ions may compete with metal ions for the cell wall ligands.

For copper ions, the pH should be lower than 5.0 to avoid precipitation of these species, as shown on copper speciation diagrams (Figure 2a). In this way, an acid treatment at pH 5.0 can allow the treated biomass to be used for metal recovery without significantly affecting biomass adsorption capacity. This fact is especially important when using a large sample amount, where the leaching and pH increase are both maximized.

3.5. Fourier transform infrared spectroscopy (FT-IR)

FTIR spectroscopy has been frequently used to detect vibration frequency changes in seaweeds[14,16,48-50]. This technique has been used to evaluate the presence of heavy metals in the biomass algae. The extension of band shifting indicates the degree of interaction of functional groups with metal cations[50].

In this study, FTIR spectroscopy was used to evaluate the changes in vibration spectra of raw and acidic algae (pH 5.0). The same evaluation was performed after copper and nickel loading on raw and acidic algae.

3.5.1. Identifying the peaks on raw algae

For raw *Sargassum filipendula* algae a broad band centered at 3430 and at 3284 cm^{-1} can be attributed to stretching modes of amino ($–NH_2$) and alcohol groups (–OH), respectively. According to Svecova et al.[51], the presence of amine groups is usually confirmed by the presence of a shoulder around 3265 cm^{-1}, this peak is frequently hidden by vibrations of –OH groups. The signal at 2928 cm^{-1} is related to C–H stretching modes, and the asymmetric stretching of carboxylate O–C–O vibration at 1640 cm^{-1}. The band at 1411 cm^{-1} may be due to C–OH deformation vibration with contribution of O–C–O symmetric stretching vibration of carboxylate group[52]. The bands at 1530-1560 cm^{-1} can be assigned to amino groups (NH stretching)[16]. A band at 1531 cm^{-1} was found for algae. The bands at about 1235cm^{-1} representing $–SO_3$ stretching are mainly present in sulfonic acids of polysaccharides, such as fucoidan[49]. The band at 1022 cm^{-1} is assigned to the C-O stretching of alcohol groups[49].

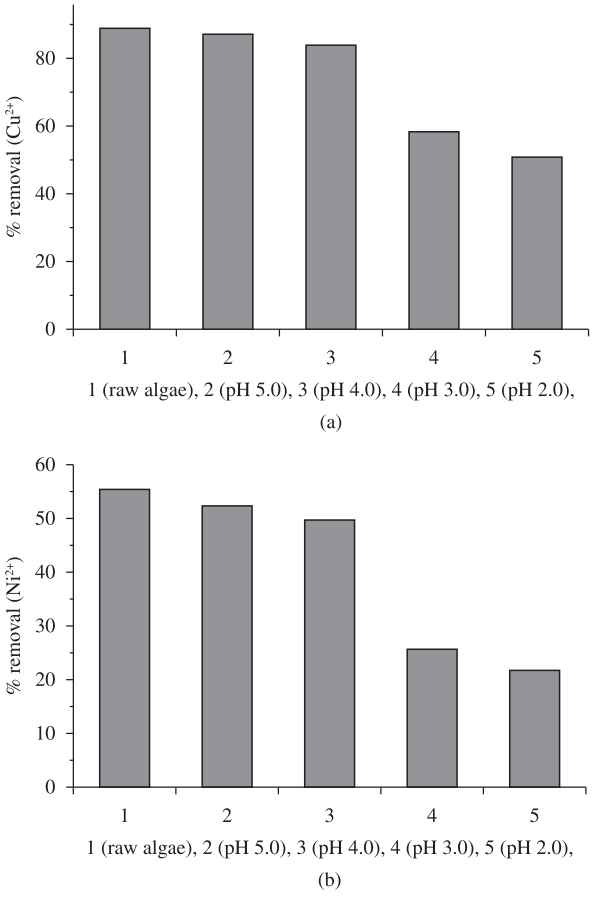


Figure 3. Comparison of a) %Cu^{2+} removal; and b) %Ni^{2+} removal of raw and acidified *Sargassum filipendula*.

3.5.2. Changes promoted by treatment acid at pH 5.0

Table 3 shows the changes observed after biomass acidification were: from 1640 to 1612 cm^{-1}, 1531 to 1533 cm^{-1}, 1411 to 1417 cm^{-1}, 1235 to 1250 cm^{-1} and 1022 to 1039 cm^{-1}. That is, the carboxylic, alcoholic, and sulfonate groups were significantly influenced by the biomass acidification.

In accordance with Sheng et al.[49], a strong peak at 1640 cm^{-1} is due to the carboxylate salt COO–M, where M can be Na^+, K^+, Ca^{2+}, and Mg^{2+} metals, which are naturally present in marine algae. This band changes to 1612 cm^{-1} after acid treatment, that is, with acid treatment metal species (M) naturally present in marine algae were replaced by hydrogen ion.

After the treatment with acid, the presence of new peaks was observed in the region of 950-750 cm^{-1}. This region is most discussed in carbohydrates[52,53]. The treatment of algae with HCl can have caused the lyses of alginate and fucoidan polysaccharide, presents in the algae.

3.5.3. Changes promoted by Cu^{2+} and Ni^{2+} loading in raw and acidified algae

Table 4 shows that after Cu^{2+} binding, in raw algae, the asymmetric carboxyl stretching band shifted from 1640 change to 1636 cm^{-1}, and after Ni^{2+} binding, it shifted to 1632 cm^{-1}. The band attributed to the symmetric stretching of these same groups at 1411 cm^{-1}, changed to 1415 cm^{-1} for Cu^{2+} and to 1370 cm^{-1} for Ni^{2+}.

According with the Table 5, for treated algae, the band at 1612 cm^{-1} changed to 1635 and 1627 cm^{-1} after copper and nickel binding, respectively, while the band at 1417 cm^{-1} moved to 1398 and 1421 cm^{-1} after copper and nickel binding, respectively.

This shift can be explained by the associations of the carbonyl groups with metal ions[49]. The large variation for copper ions can be attributed to the largest affinity of these species and the algae.

Raize et al.,[18] observed that during the biosorption process the metal ions in solution can also be exchanged with biomass protons. Copper biosorption with *F. vesiculous* was estimated to be 77% due to ion exchange with Ca^{2+}, Mg^{2+}, Na^+ and K^+ ions[54].

Table 3. Changes promoted by treatment acid at pH 5.0

Raw algae cm^{-1}	Acidified algae (pH 5.0) cm^{-1}
2928	2950
1640	1612
1531	1533
1411	1417
1235	1250
1022	1039

Table 4. Changes promoted by Cu^{2+} and Ni^{2+} loading in raw algae.

Raw algae cm^{-1}	In nature algae saturated with Cu^{2+} cm^{-1}	In nature algae saturated with Ni^{2+} cm^{-1}
2928	2943	2921
1640	1636	1632
1531	-	1535
1411	1415	1370
1235	1220	1213
1022	1037	1033

Studies using *Sargassum* stated that cadmium biosorption occurred by the formation of ionic bridges between the metal and two carboxyl groups or a bidentate chelating complex with one carboxyl group[19].

Regions of the alginate polymer that are rich in "G" residue, provide a multi-dentate environment for complexation, whereas in regions that are rich in mannuronic acid, complexation would be predominantly monodentate and therefore, weaker. In guluronic acid, the ring oxygen and the axial O-1 form a spatially favorable environment with –COO, as opposed to the equatorial O-1 which occurs in mannuronic acid residues[1,38].

The analysis of spectra obtained before and after copper and nickel binding indicated that the –NH group was involved in biosorption process. For raw algae, the band at 1531 cm^{-1} disappears after copper adsorption and moves to 1535 cm^{-1} after nickel adsorption, while for treated algae the band at 1533 cm^{-1} changes to 1543 cm^{-1} after copper adsorption. The changes in intensity of the bands in the 3430 cm^{-1} region also suggested changes in the amino groups present in the biomass.

The infrared frequency at 1235 cm^{-1} represents SO_3 stretching. For raw algae, Table 4, this band changes to 1220 and 1213 cm^{-1} in the presence of Cu^{2+} and Ni^{2+} respectively. Analyzing Table 5 for the band at 1250 cm^{-1} (acidic algae), nearly the same frequency was observed for Ni binding (1248 cm^{-1}), while for Cu binding, it changed to 1226 cm^{-1}. This fact can be explained by copper complexation on fucoidan groups present in the algae, either for raw or acid algae. For nickel ions, after acid treatment, metal complexation was not observed.

Table 5. Changes promoted by Cu^{2+} and Ni^{2+} loading in acidified algae.

Acidified algae (pH 5.0) cm^{-1}	Acidified algae saturated with Cu^{2+} cm^{-1}	Acidified algae saturated with Ni^{2+} cm^{-1}
2950	2945	2938
1612	1635	1627
1533	1543	-
1417	1398	1421
1250	1226	1248
1039	1037	1036
824	822	852
740	743	-

The band at 1022 cm^{-1} was assigned to the C-O stretching of hydroxyl groups. In raw algae, this band shifts to 1037 and 1033 cm^{-1} after Cu^{2+} and Ni^{2+} sorption, respectively.

According to Mackie[55], alginates showed two characteristic bands at 808 and 787 cm^{-1} in the IR spectra, assigned to mannuronic and guluronic acids, respectively. Here, the band identified at 824 cm^{-1} (Table 5) moved to 822 cm^{-1} after Cu^{2+} binding and to 852 cm^{-1} in the presence of Ni^{2+}. After Cu^{2+} sorption the band at 740 cm^{-1} slightly moved to 743 cm^{-1}.

3.6. SEM/EDX - Sargassum filipendula

The morphology of algae surface was analyzed by scanning electron microscopy before and after the acidic treatment and after copper and nickel loading. The presence of diatom shells in the outer algae surface was systematically observed. In diatoms, the cell wall is composed of silica, to which protein and polysaccharide are added. Even after the diatom dies and the organic materials have disappeared, the external structure remains, showing that the siliceous component is indeed to decay of these diatom frustules, they remain intact for long periods of time and constitute some of the best algal fossils ever found[56]. Diatoms outer shells are fixed on algae surface. There was a high variability of these shell quantity, with regions with large amounts and regions where diatoms are not observed, (Figure 4a-e).

Figure 4(b-e) shows the effect of treatment with acid. These changes were probably caused due to strong cross-linking between the H^+ and negatively charged chemical groups in the cell polymer. In the raw algae are high concentrations of calcium, sodium, magnesium, etc. (Table 6) and these bind to alginic acid monomers. This binding creates a net of cross-linking.

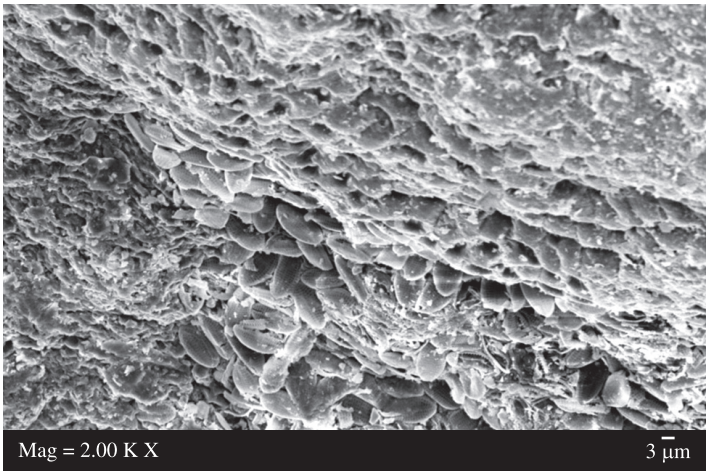
When the *Sargassum filipendula* samples were exposed to heavy metal solution, the cations replaced some of the cation initially present in the cell wall matrix and created stronger cross-linking[18].

In order to characterize the chemical composition and the location of the diatom outer shell on algae surface scanning electron microscopy (SEM) with energy dispersive X-ray spectroscopy (EDX) was used, Figures 5a and b. High silicon amounts were observed in the regions where the diatom outer shells were present, due to the fact that silicate groups are the principal components of these microorganisms.

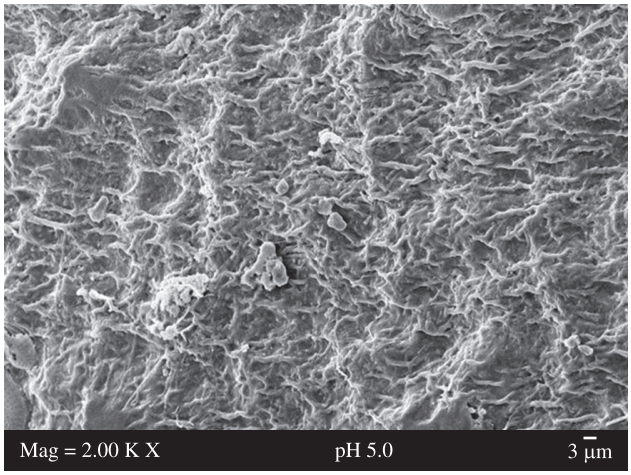
The Figure 6 and 7, presents scanning electron microscopy (SEM) with energy dispersive X-ray spectroscopy (EDX), when the algae (treated pH 5.0) it is saturated with the heavy metals copper and nickel, respectively. Again the presence of diatoms is observed and the quantity of metals copper and nickel did not suffer variation in locations with or without the presence of diatoms.

Table 6. Elemental composition of *Sargassum filipendula* treated in the different pHs.

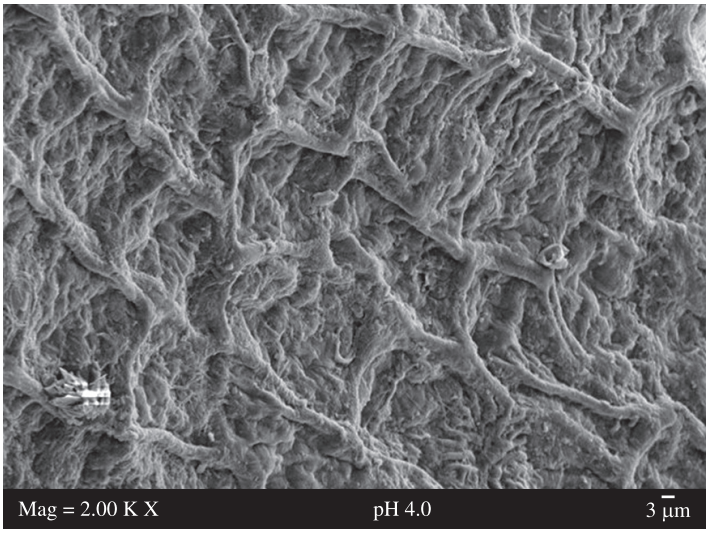
Element	Composition									
	In nature		pH 5.0		pH 4.0		pH 3.0		pH 2.0	
	wt. (%)	at. (%)	wt. (%)	at. (%)	wt. (%)	at. (%)	wt. (%)	at. (%)	wt. (%)	at. (%)
Mg	1.43	13.07	1.49	17.56	0.72	10.31	0.52	8.52	0.22	5.09
Na	1.09	10.49	-	-	-	-	-	-	-	-
Al	1.58	13.02	0.61	6.49	0.89	11.44	0.63	9.19	0.21	4.38
Si	4.11	32.56	1.71	17.49	1.93	23.73	1.57	22.15	0.61	12.29
Cl	0.61	3.82	-	-	0.24	2.37	0.35	3.89	1.35	21.73
Fe	0.85	3.37	0.34	1.75	0.58	3.57	0.38	2.69	-	-
S	-	-	3.06	27.33	1.63	17.63	2.13	26.25	2.29	40.71
K	2.31	13.13	0.61	4.45	0.33	2.89	0.34	3.39	-	-
Ca	1.91	10.57	3.49	24.93	3.25	28.05	2.43	23.92	1.11	15.79
Total	13.88	100.00	11.31	100.00	9.57	100.00	8.35	100.00	5.79	100.00



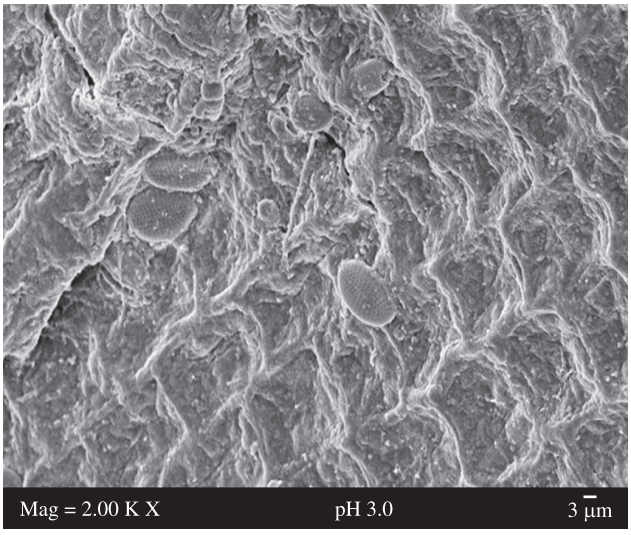
(a)



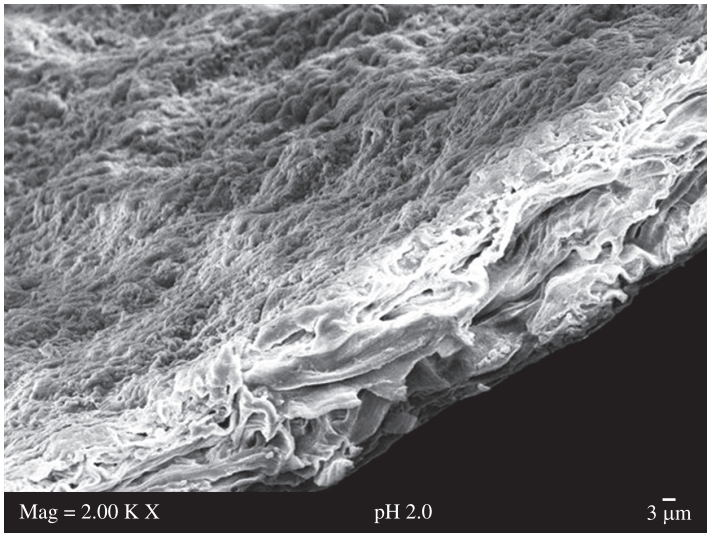
(b)



(c)



(d)



(e)

Figure 4. Scanning electron microscopy (SEM) micrographs (2000× magnification) of a) raw *Sargassum filipendula*; b) acid treatment *Sargassum filipendula* pH 5.0; c) acid treatment *Sargassum filipendula* pH 4.0; d) acid treatment *Sargassum filipendula* pH 3.0; and e) acid treatment *Sargassum filipendula* pH 2.0.

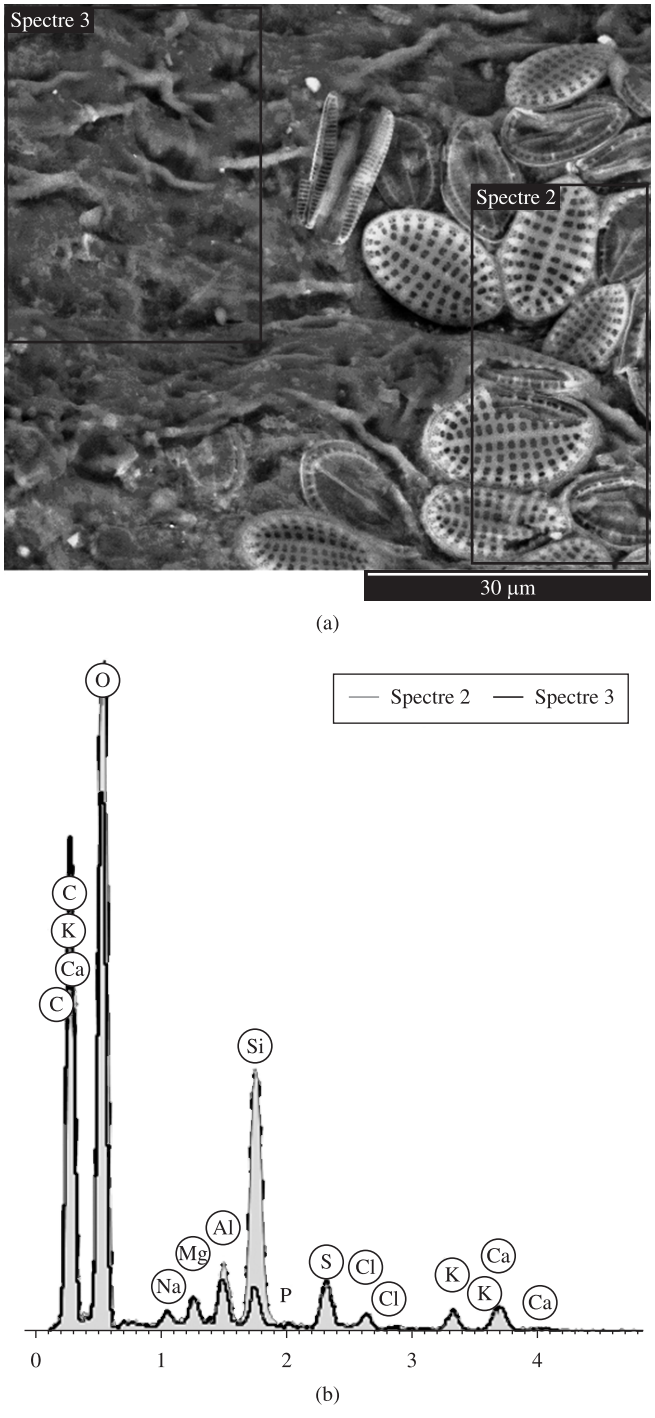


Figure 5. a) Scanning electron microscopy (SEM) micrographs of *Sargassum filipendula* treated pH 5.0; and b) energy dispersive X-ray spectroscopy (EDX) in the regions of spectra 2 and 3 of the SEM.

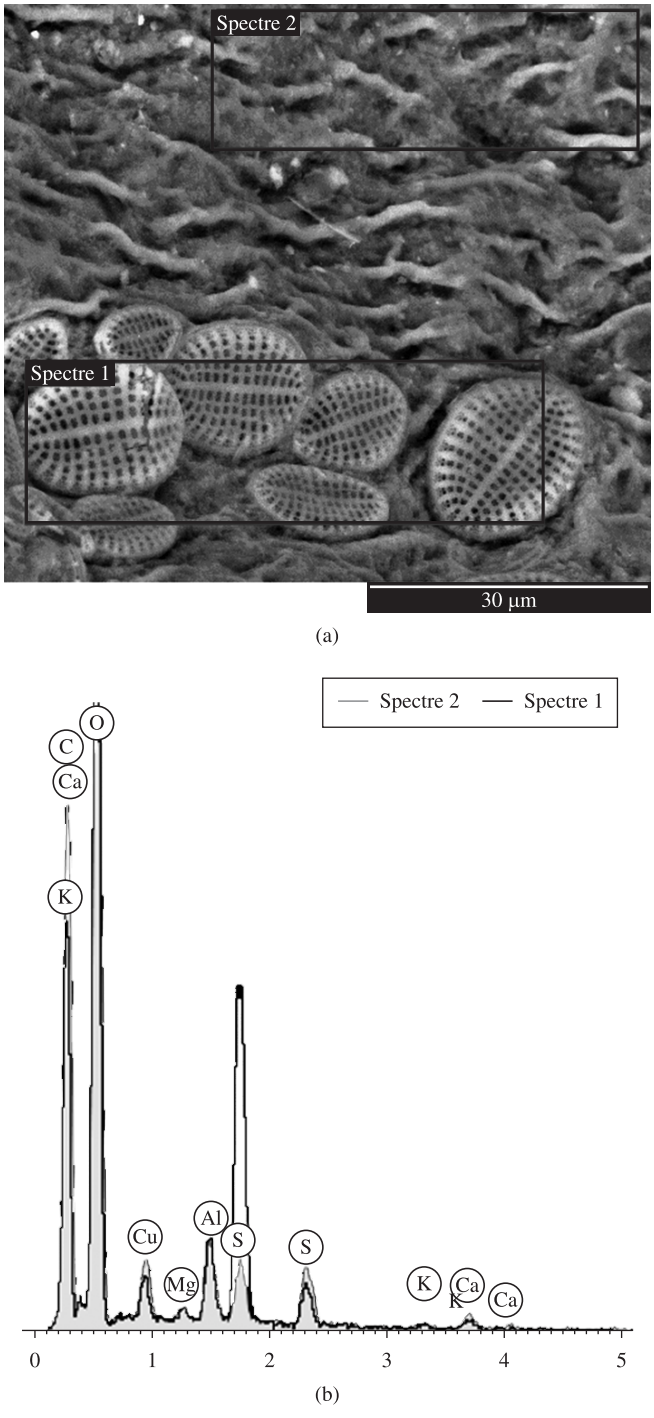
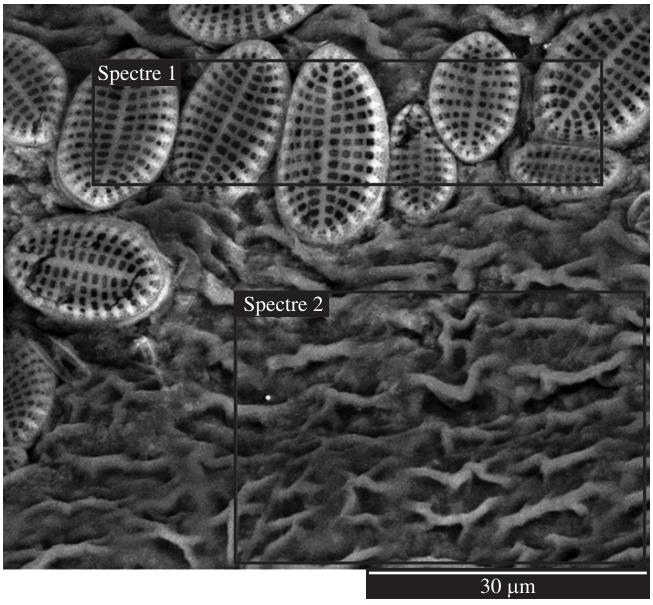


Figure 6. a) Scanning electron microscopy (SEM) micrographs of *Sargassum filipendula* treated pH 5.0 saturated with copper; and b) energy dispersive X-ray spectroscopy (EDX) in the regions of spectra 2 and 1 of the SEM.

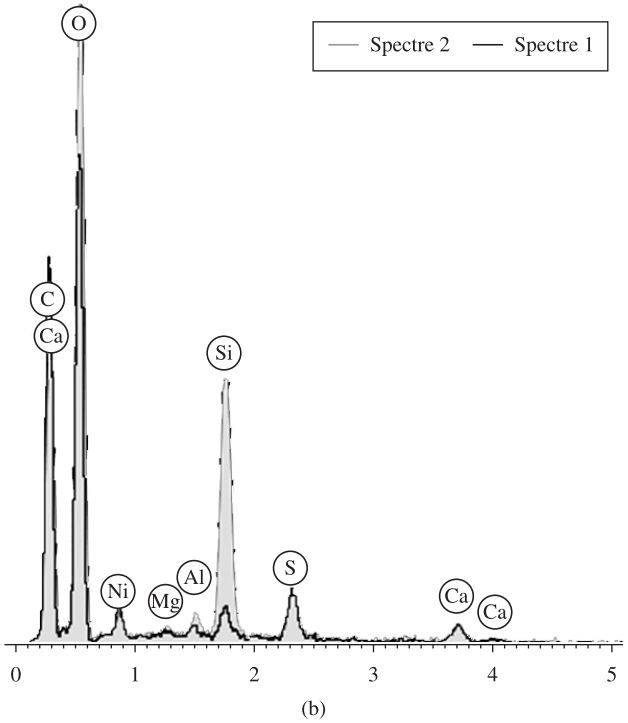
The cell walls of brown algae generally contain three components: cellulose, the structural support; alginic acid, a polymer of mannuronic and guluronic acids and the corresponding salts of sodium, potassium, magnesium and calcium; and sulfated polysaccharides (fucoidan matrix)[56].

Table 6 shows qualitative results of chemical distribution on algae surface at different pHs. The objective is to compare between the chemical distributions of metals at different pHs for all samples. The chemical quantification was performed in regions where diatom outer shells were present. It was observed that the acid treatment resulted in the removal of metals such as: Mg, Na, Al, Si, Ca, Fe and K from algae biomass.

This fact can be related to FTIR results, where the absorption in 1640 cm^{-1} represents the carboxylate salt COO-M, where M can be the naturally found metals in the algae. The spectra showed alterations after the acid treatment and that was justified by the elimination of these ions. This behavior was more significant in algae acidified in smaller pH values.



(a)



(b)

Figure 7. a) Scanning electron microscopy (SEM) micrographs of *Sargassum filipendula* treated pH 5.0 saturated with nickel; and b) energy dispersive X-ray spectroscopy (EDX) in the regions of spectra 1 and 2 of the SEM.

4. Conclusions

In this work the characterization and evaluation of copper and nickel biosorption was studied on acidic *Sargassum filipendula* algae. The following conclusions were drawn from the study:

- A controlled acid treatment, at the appropriate pH (pH 5.0), prevents the release of organic material during metal sorption and keeps the sorption capacities in the same order of magnitude of raw algal material. Additionally, this pre-treatment allows limiting pH variation during the sorption process (especially for column application).
- Depending on the algal species used for the extraction of alginic acid its M/G ratio presents great variations. In this work the M/G ratio found was 0.50.
- The Fourier transform infrared (FT-IR) analysis demonstrates similar chelating characteristics of copper and nickel coordination to the functional groups in the cell wall of *Sargassum filipendula* algae. The presence and participation of carboxylic and alcoholic groups in the alginate biopolymer, sulfonate group in the sulfate biopolymer (fucoidan) and amino groups in amino acids are responsible for copper and nickel adsorption.
- The morphology of algae surface was analyzed by scanning electron microscopy and the presence of diatom outer shells was observed, though the presence of this diatom did not have a significant impact in copper and nickel binding.

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