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Arsenic Sorption on Chitosan-Based Sorbents: Comparison of the Effect of Molybdate and Tungstate Loading on As(V) Sorption Properties

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Abstract

Modified chitosan gel beads, prepared by molybdate and tungstate coagulation methods, were tested for As(V) removal from solutions in the range of 5–200 mg As L⁻¹. The sorbent is efficient at removing As(V) from acid solutions (optimum pH close to 3), the sorption capacities for As uptake in molybdate- and tungstate-loaded beads are 75 and 44 mg As g⁻¹ of dry mass, respectively. The mechanism of As(V) sorption is related to the ability of molybdate and tungstate ions to complex As(V) ions in acid solutions. As(V) sorption process is mainly influenced by the presence of phosphate ions, but there is no influence of co-ions as nitrate and chloride. Arsenic desorption can be performed using phosphoric acid solutions. Arsenic adsorption proceeds in acidic solutions with a partial release of molybdate and tungstate and with residual concentrations of arsenic above the regulations for drinking water. For that reason, this material is a candidate for the treatment of industrial effluents.

Graphic Abstract

Keywords Arsenic · Treatment · Chitosan beads · Coagulation

Introduction

In the environment, arsenic exists in various forms. In natural waters, the two most common are arsenites (AsO_3^{3-}) and arsenates (AsO_4^{3-}), known as As(III) and As(V), respectively. Out of these, the trivalent form is more toxic and difficult to remove [1]. Although there is pollution by arsenic from anthropogenic sources, most of the arsenic present in groundwater has a geological origin mainly associated with an oxidation reaction of sulfide minerals in metasedimentary rocks [2]. In some countries (e.g., India, Bangladesh, and the United States) severe adverse health effects have been reported [3], because of arsenic naturally-occur in water for human consumption. Arsenic exposure from water and food associated with carcinogenic effects has been a growing concern in recent decades, as well as neurological, cardiovascular, respiratory, and liver damage, among others, skin, bladder, and lung cancer have been documented [4, 5]. Therefore, the presence of arsenic in natural waters is a global problem, and its removal is of vital importance.

The use of biomaterials for the treatment of water containing heavy metals has gained attention due to their simplicity and effectiveness, as well as their economic advantages [6]. Among the biosorbents used, chitosan (CTS) it is considered as an ideal candidate in the removal of heavy

metals from wastewater, due to its amine groups that are strongly reactive with metal ions and are responsible for the uptake of metal cations by chelation. The amino groups easily protonate in acidic solutions that may cause electrostatic attraction of anionic compounds [7]. In addition to its chemical characteristics, chitosan has the advantage of being highly biocompatible and biodegradable [8]. However, although chitosan has been used successfully for the removal of metals such as Cr, Pb and Cu [9–12] it has a significantly lower sorption capacity for As(V), which in most cases does not exceed 14 mg As g⁻¹ and it is even lower for As(III). In such a case, it is necessary to chemically modify the biopolymer to increase its sorption capacity by incorporating other functional groups with a strong affinity for the As. Molybdate can complex arsenate ions, thus modifying chitosan with molybdate may increase the arsenic sorption capacity of chitosan. There is a previous work that prepared biopolymer beads either impregnated with molybdate [13] or coagulated into a molybdate bath, that reached a removal capacity as high as 230 mg As g⁻¹ Mo [14]. One objective of this work is to prepare smaller beads than those previously reported using an encapsulator.

Moreover, we are interested in preparing modified chitosan beads with tungstate since it is an ion that shares many chemical properties with molybdate. To our knowledge,

there are no previous studies on arsenic sorption using chitosan modified with tungstate. The aqueous chemistry of tungsten compared with molybdate is an advantage because hexavalent tungsten form is very stable in most hydrothermal fluids [15]. For these reasons, in this work, we evaluate As(V) sorption capacity of CTS beads modified with either molybdate or tungstate using sorption isotherms and the effect of the drying process on sorption kinetics.

Materials and Methods

Materials

Chitosan medium molecular weight and deacetylation degree of 75% was purchased from Sigma Aldrich (Batch# STBF3507V). Molecular weight was determined by measuring the intrinsic viscosity ($M_w = 125,000 \text{ g mol}^{-1}$) using a Cannon–Fenske capillary tube with the Mark–Houwink constants proposed by Rinaudo in 2006 [16] to avoid aggregate formation [17]. Reagents (analytical grade): $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24} \cdot 4\text{H}_2\text{O}$ and $\text{Na}_2\text{HAsO}_4 \cdot 7\text{H}_2\text{O}$ were purchased from Fluka AG and Merck respectively, and $\text{Na}_2\text{WO}_4 \cdot 2\text{H}_2\text{O}$ from Carlo Erba.

Preparation of Molybdate-Loaded and Tungstate-Loaded Chitosan Beads (CTS–Mo and CTS–W)

Chitosan (1.5% w/w) was dissolved in an acetic acid solution (4% v/v). The viscous solution was dropped using an encapsulator (BUCHI B-390) into ammonium heptamolybdate and sodium tungstate solutions at a concentration of 7 g L^{-1} of Mo and W, respectively. Coagulation is evaluated at three different pH values adjusting it with HCl and NaOH. Beads remained in the coagulation bath for 24 h and were rinsed with distilled water until neutral pH, then oven-dried at 50°C until constant weight. Before carrying out the drying process, part of the beads was placed in a 0.1 M phosphoric acid solution for 24 h to remove the labile part of the metal. For kinetic experiments, some beads were freeze-dried, some others were oven-dried, and the rest were kept hydrated.

Characterization

SEM–EDX SEM and SEM–EDX analysis were performed in oven-dried and freeze-dried beads, using an environmental scanning electron microscope Quanta FEG 200. The total concentration of Mo and W in the gel phase was determined using ICP–AES analysis (ICP–AES JY Activa M) by mineralization of the beads. For this purpose, we put 100 mg

of wet beads into a 50 mL beaker and added 3.0 mL of hydrogen peroxide (30% v/v) and 1.0 mL of nitric acid. The beaker was heated at 80°C in a thermostated bath around 30 min until the beads dissolve completely. After cooling, the solution was transferred into a volumetric flask and averaged to 25 mL, a sample of this solution was filtered through a Whatman filter membrane (pore size $1.2 \mu\text{m}$) for ICP–AES analysis. The dry mass was obtained by weight loss at 50°C for at least 72 h. Experiments were run in triplicate.

Specific Surface Area (SSA) An analysis of the specific surface area was performed according to the standard gas adsorption method and the application of the BET equation [18], using nitrogen at 77 K. The measurement was performed using a ChemBET TPR/TPD Chemisorption analyzer, with 50 mg of sample for CTS–Mo and CTS–W beads, oven-dried, and freeze-dried. Before analysis, the samples were degassed for 5 h at 200°C , and the SSA of each sample was calculated using the single point method at a relative pressure (P/P_0) of 0.2.

FTIR Infrared analysis is carried out to clarify the interaction mechanism between Mo or W with chitosan and As(V). FTIR analysis was performed on CTS beads coagulated in a NaOH bath as well as on CTS–W and CTS–Mo beads coagulated by the ionotropic gelation method. The CTS–W and CTS–Mo beads were analyzed before and after arsenic sorption. For this procedure, a fraction of the corresponding oven-dried beads were ground and analyzed using an ATR accessory on a Thermo Scientific Nicolet iS5 from 4000 to 400 cm^{-1} with a resolution of 4 cm^{-1} and 25 scans.

Arsenate Sorption and Desorption

Previous studies determined that the optimum pH for arsenate sorption is around 3 [14]; this pH was fixed at the beginning of the process and was measured again at the end. For constructing the sorption isotherms, 20 mg of oven-dried beads were put into a 20 mL of As(V) solution at different concentrations (from 5 to 200 mg L^{-1}) at 20°C for at least 96 h. Subsequently, the concentration of As was measured by ICP–AES after filtering the sample through a Whatman filter membrane (pore size $1.2 \mu\text{m}$). For kinetic studies, 400 mg of beads (hydrated, oven-dried, and freeze-dried) were placed in 500 mL of As(V) solution, then samples of the solution were withdrawn at fixed times and filtered before analysis.

In this study, arsenic desorption is performed using phosphoric acid. As previous investigations have shown, phosphate solutions are efficient in arsenic removal [13]. For this purpose, 40 mg of oven-dried beads of known arsenate concentration was brought into contact with 20 mL of eluent at a concentration of 0.1 M for 24 h. Concentrations of As, Mo, and W in solutions were determined by ICP–AES analysis.

Interference of Co-ions and Competitor Ions

The effect of different ions on arsenic sorption is tested using phosphate, sulfate, nitrate, and chloride solutions prepared at molar ion:As ratios corresponding to 1:1, 10:1, and 100:1 with a sorbent dosage of 1.0 g L^{-1} . Also, considering the actual composition of contaminated effluents [19, 20], solutions were prepared to reproduce these conditions and tested to evaluate the effect of different ions present during the sorption process. For these tests, the sorbent dosage is 1.25 g L^{-1} .

Results

Sorbent Manufacturing—Influence of the pH of the Coagulating Bath on Metal (Mo or W) Content

Figure 1 shows the variation of the total Mo and W contents per gram of dry mass in the chitosan beads as a function of the initial pH. For the initial tests, the pH was set at the beginning, and no considerable variation was observed due to the short time. However, in the manufacture of the subsequent batches, the pH was controlled during the process. The initial metal concentration in solution was set at 7 g L^{-1} for both molybdate and tungstate [14]. For molybdate, the maximum loading of $3.31 \text{ mmol Mo per g}$ of dry mass is reached at pH 6, which is very close to the value at pH 5. According to the distribution diagram (Fig. SM1a of the supplementary material), calculated using Medusa software, the polynuclear hydrolyzed molybdate species present are $\text{Mo}_7\text{O}_{23}(\text{OH})^{5-}$ and $\text{Mo}_7\text{O}_{24}^{6-}$ in near-equivalent proportions. These two species represent more than 90% of the total molybdenum. The strong affinity of chitosan for molybdate has been directly correlated to the binding of CTS with this polynuclear polyhydrolyzed species [21]; this is consistent with a lower quantity of molybdate bound to chitosan at the

Fig. 1 Variation of the total amount of molybdate and tungstate in chitosan beads as a function of the initial pH. C_0 : $7 \text{ g Mo or W L}^{-1}$

initial pH near 7, which is $2.52 \text{ mmol Mo g}^{-1}$. The process allows incorporating comparable amounts of tungsten: at pH 8, the W loading approaches $3.12 \text{ mmol W g}^{-1}$; this is slightly higher than the levels reached at pH 6. Again, for tungstate, the decrease of the fraction of polynuclear species (i.e., the predominance of the simple oxo complex WO_4^{2-}) corresponds to a progressive diminution of tungsten loading (Fig. SM1b of the supplementary material). In 2003, Gustafsson reported that the sorption of tungstate onto ferrihydrite was stronger than the accumulation of molybdate, despite very similar chemical characteristics and comparable pK_a values [22]. This behavior was also reported by Xu et al. in 2006 [23] for the comparative sorption of Mo and W onto goethite.

The literature provides limited information regarding the tungstate-chitosan interaction, but tungsten sorption depends to a large extent on its aqueous chemistry. The most stable oxidation state of tungsten is 6+, which forms some oxo complexes (e.g., mono-tungstate WO_4^{2-} , para-tungstate A $\text{W}_7\text{O}_{24}^{6-}$ and para-tungstate B $\text{H}_2\text{W}_{12}\text{O}_{42}^{10-}$). Generally, non-protonated monomeric species (WO_4^{2-}) are the most stable under neutral to alkaline conditions because acidic pH promotes polymerization leading to the formation of polytungstate species [24, 25]. At pH 8, the predominant species is the polytungstate $\text{W}_6\text{O}_{21}^{6-}$. Gecol et al. in 2006 [26] suggested that an electrostatic mechanism is responsible for tungstate binding on chitosan: in acidic solutions, the protonation of amine groups enhances the attraction of polynuclear anionic species (anionic polytungstate species). In this case, when working at pH 6, the amount of W loaded ($3.02 \text{ mmol W g}^{-1}$) has little variation compared to the amount loaded at pH 8. However, as the pH of the coagulation bath decreases, it becomes more difficult to form the beads due to the dissolution of chitosan in the acidic medium.

Chemical Stability of Beads

Preliminary studies and previous work have pointed out the presence of traces of free, absorbed, or weakly bound molybdate in the polymer network. This little fraction is relatively mobile, and its release into the solution may cause the complexation of As(V), which, in turn, may be less available for binding on the composite sorbent. Preliminary tests on these sorbents have shown that a fraction of arsenic remained unavailable to the sorbents, consistently with the previous comment. A dilute solution of phosphoric acid for removing this labile fraction of molybdate (and tungstate) from polymer network is used because of the affinity of phosphoric acid for desorbing molybdate (and tungstate) [13]. Figure 2 shows the amount of molybdate and tungstate loaded into the beads before and after washing with a solution of phosphoric acid

Fig. 2 Variation of the total amount of **a** molybdate (pH 7, C_0 : 7 g Mo L⁻¹) and **b** tungstate (pH 8, C_0 : 7 g W L⁻¹) content in two different batches of chitosan beads, before and after treatment with phosphoric acid 0.1 M

0.1 M, for two different batches. As expected, in both cases, the metal loading in the beads decreases after the treatment with phosphoric acid: for molybdate-loaded sorbent, the average value decreases 12%, while for tungstate-loaded the metal loading decreases only 5%. Phosphate, molybdate, and tungstates are tetrahedral oxyanions that strongly bind adsorbates that share similar characteristics in terms of ion-proton sorption; thus, they compete for the adsorption sites [27]. However, the most significant decrease in molybdenum content could be attributed to the formation of phosphomolybdate species; indeed, phosphate can react with molybdate when present at high concentration [23].

SEM–EDX Characterization of Beads

The type of drying may drastically change the structure of the beads. Figure 3 compares the environmental scanning electron micrographs of the CTS–Mo and CTS–W beads after oven-drying and freeze-drying processes. Molybdate and tungstate beads that were freeze-dried exhibit a highly porous structure, formed by the voids left by the removed solvent, due to the fact that the sample should be kept below the glass transition temperature when removing the frozen solvent under vacuum; the solvent then acts as a porogen to produce this type of structure in materials, which is well documented [28]. On the other hand, the oven-drying technique leads to the formation of highly compact structures due to the elimination of water molecules during the process increases the concentration of the solvent, creating a fluid drag, the particles then tend to form aggregates of higher stability [29]. The oven-drying does not allow managing capillary forces during the drying process as it may occur with freeze-drying and even better with drying under supercritical CO₂ conditions [30, 31]. The drying process affects the diameter of the beads: for CTS–Mo beads, it changes from 1.02 ± 0.11 mm when freeze-dried to 0.44 ± 0.06 mm when oven-dried. Similar changes are for

CTS–W beads: their diameter decreases from 1.06 ± 0.12 mm to 0.46 ± 0.07 mm, respectively. The distribution of molybdenum and arsenic is roughly homogeneous (Fig. SM2 of the supplementary material); this means that chitosan ionotropic gelation with molybdenum is occurring in the whole mass of the beads and that arsenic is spread in the entire volume (i.e., all molybdate reactive groups are accessible; no concentration gradient was detected). The freeze-drying process allows the formation of a wide-open structure compared to a dense material formed in the oven-drying process; this is expected to influence mass transfer properties, diffusion, and uptake kinetics.

Specific Surface Area

The results of SSA are shown in Table 1, as expected, they are consistent with SEM analysis. The morphology of the oven-dried beads leads to the loss of pores in the material, thus obtaining a very low surface area; this is consistent with the time required for this material to reach the sorption equilibrium. The surface area of the CTS–W oven-dried beads is the lowest with a value of less than 1.0 m². When the surface area is relatively small, (NIST considers < 2 m² g⁻¹) [32] it is recommended to use krypton instead of nitrogen; however, the equipment manufacturer reports that it is possible to measure absolute surface areas as low as 0.5–1.0 m² using nitrogen as adsorption gas [33]. On the other hand, freeze-dried beads retain the porous structure with a surface area 20 times greater than that of oven-dried beads, which implies an adsorption process much faster at the beginning.

FTIR Characterization for Beads Interactions

Interaction Between Chitosan and Mo or W Groups Metal speciation controls molybdate/chitosan interactions and affects the predominance of polynuclear anionic species

Fig. 3 SEM micrographs and EDX element cartographies of CTS–Mo and CTS–W beads, oven-dried and freeze-dried

Table 1 Specific surface area for CTS–Mo and CTS–W beads, freeze and oven-dried

Sorbent	Specific surface area, m ² g ^{−1}	
	Oven-dried	Freeze-dried
CTS–Mo	1	24
CTS–W	< 1	22

[21]. Heptamolybdate species have a high anionic charge, which allows polyoxyanions to react with the protonated amino groups of the same or different chitosan chains. Also, the presence of a large number of oxygen atoms and -OH groups allow the formation of additional hydrogen bonds. Thus, multiple reactions lead to strengthening the structure of the beads [13]. Tungstate ions may behave similar to molybdate ions, due to the type of polynuclear anionic species present in the coagulation bath. Figure 4 shows the FTIR spectra of three samples: CTS beads coagulated by the traditional method (NaOH bath), and CTS–Mo and CTS–W beads formed by the ionotropic gelation method in Mo and W baths, respectively. The CTS beads spectrum (black) exhibits the characteristic bands of the CTS biopolymer. In the region of 3600 to 3000 cm^{−1}, the stretch bands of

Fig. 4 FTIR spectra of CTS, CTS–Mo, and CTS–W beads

the amino-NH₂ and hydroxyl-OH groups appear as a broad peak due to the multiple hydrogen bonding. The bands at 2875–2930 cm^{−1} are due to stretching of –CH groups. The amide I and amide III bands confirm the presence of residual N-acetyl groups at 1645 cm^{−1} for C=O stretching vibration

(amide I band) and at 1315 cm^{-1} for the stretching of C–N group (amide III). The band at 1538 cm^{-1} is due to the NH bending. At 1403 cm^{-1} appears the vibration of OH group in the ring and at 1370 cm^{-1} , the vibration of CH_3 of the amide group. The bands around 1057 and 1030 cm^{-1} correspond to the C–O–C stretching of the glycosidic linkage. Finally, the low-intensity bands at 850 and 838 cm^{-1} are due to the vibration of the CH_3COH group [34, 35].

Concerning the oxyanions of Mo and W, the region of the appearance of Mo–O and W–O vibration bands, is practically the same for both metals [36]. The adsorption bands for Mo are reported in the region from 1000 – 700 cm^{-1} [37] and the stretching W–O between 880 – 820 cm^{-1} [38]. The CTS–Mo beads spectrum (red) shows a peak at 880 cm^{-1} of medium intensity and a shoulder near 840 cm^{-1} . The band at 850 cm^{-1} shifts to 880 cm^{-1} and increases its intensity, due to an interaction between Mo–O and oxygen of OH groups of CTS; because the 880 cm^{-1} band also corresponds to the Mo–O–Mo bonds of the polynuclear species. These results are consistent with the mechanism proposed by Bertoni et al. in 2018 [39], where the hydroxyl groups of chitosan are the active binding sites to coordinate molybdate anions. According to their results, the protonated amino groups stabilize the negative charges of heptamolybdate anions, which in addition to the binding with hydroxyl groups, results in the retention of molybdate.

The CTS–W beads spectrum (blue) shows two defined peaks at 875 and 835 cm^{-1} , which indicates an interaction between W–O and OH groups of CTS. Besides, the band at 1538 cm^{-1} of NH bending of CTS shifts to 1520 cm^{-1} for both CTS–Mo and CTS–W beads; this is a characteristic band of the interaction of an amide II in CTS with metallic ions [37].

Interaction Between Mo or W and As(V) Dambies et al. [13] suggested the mechanism of adsorption of As(V) onto chitosan beads impregnated with molybdate. The arsenate complex with molybdate ions due to the formation of the molybdoarsenate complex. Later, in 2002, they confirmed by an XPS analysis that arsenic sorption could only occur in molybdate since the adsorption capacity of As(V) decreases if molybdate is chemically reduced [14].

Our FT-IR analysis (Fig. 5) performed on chitosan beads loaded with Mo and W before and after arsenic sorption confirms the formation of the molybdoarsenate complex with the disappearance of the 880 cm^{-1} band corresponding to the Mo–O asymmetric vibration and the appearance of a new characteristic peak of arsenate adsorption at 836 cm^{-1} due to the stretching vibration of As–O [40]. On the other hand, typical bands for the oxo complexes of W and As appear between 836 and 856 cm^{-1} due to the deformation of the AsO_4 tetrahedron and bands between 682 and 783 cm^{-1} assigned to W–O–W and WO–As bonds [41].

Fig. 5 FTIR spectra of CTS–Mo, and CTS–W beads, before and after As(V) sorption

Taking into account the results obtained from FTIR analysis in this work and the studies previously mentioned, we consider that the mechanism showed in Fig. 6 could describe the interaction between chitosan, molybdate, and arsenate, and we could expect, as has been discussed, a similar behavior with tungstate ions.

Influence of pH in As (V) Sorption

Figure 7 shows the influence of pH on As (V) sorption using oven-dried CTS–Mo and CTS–W beads. Plots represent arsenic sorption as a function of Mo and W mass accumulated in the beads, as well as the percentage released in the solution. The redox potential and pH are the most important factors that control arsenic speciation, under oxidizing conditions, and at pH below 6.9, H_2AsO_4^- is the dominant species [42]. This species predominates at pH 2.5 and 4 with 75% and 99%, respectively (Fig. SM3). Although the chemistry of molybdo-arsenic anions is complicated, a mechanism similar to complexation with arsenate ions has been established [13]. In the case of tungsten, very little information is available on the interaction of its anions with arsenic, although it is known that compounds such as WAs_2 and WAs_3 exist in the tungsten-arsenic system [43]. For both sorbents, the optimum pH range turns out to be very similar: the best sorption occurs at pH close to 3. It is noteworthy that this pH also corresponds to the lowest release of active metal ions (i.e., molybdate or tungstate). The low release of coordinating metal limits the competitive complexation of arsenic with metal ions for binding on reactive groups on the sorbents, so the sorption is enhanced.

Fig. 6 **a** Possible mechanism for molybdate binding to chitosan (adapted from [39] with permission from ELSEVIER), **b** Possible structure of the arsenomolybdate complex (reproduced from [40] with permission from The Royal Society of Chemistry)

Fig. 7 Influence of pH on arsenic sorption for **a** CTS-Mo and **b** CTS-W beads. Sorbent dosage: 1.25 mg mL^{-1} , C_0 : 100 mg As L^{-1} , contact time: 72 h, T: 20°C (closed symbols: As sorption, open symbols: Mo and W release)

On the other hand, minimizing the release of molybdate and tungstate reduces the post contamination of the solution: substituting arsenic contamination by Mo or W contamination would not make sense. Above this pH, the arsenic sorption begins to decrease, and the release of molybdenum and tungsten in the beads increases. At acidic pH (around pH 1), the intense competition of protons and the speciation of arsenic are less favorable for metal binding; the sorption capacity sharply decreases.

As (V) Sorption Isotherms

Figure 8 shows a comparison between sorption isotherms of As (V) in CTS-Mo and CTS-W beads at pH 3, before and after treatment with phosphoric acid. As can be observed for both materials, sorption capacity is reduced after acid treatment, going from 1 to $0.82 \text{ mmol As g}^{-1}$ of dry mass (i.e., 116 to $108 \text{ mmol As g}^{-1} \text{ Mo}$) for CTS-Mo

Fig. 8 Influence of the phosphoric acid treatment in CTS-Mo and CTS-W beads. **a** beads without treatment **b** beads treated with phosphoric acid. Sorbent dosage: 1 mg mL⁻¹, pH: 3, contact time: 140 h, T: 20 °C

beads and from 0.59 to 0.19 mmol As g⁻¹ of dry mass (i.e., 60.8 to 21.6 mmol As g⁻¹ W) for CTS-W beads. This loss of capacity can be attributed to the fact that during treatment with phosphoric acid, not only the labile part of the metal is removed, but also phosphate anions could be bound to the sorbent surface. In this case, phosphate anions would compete directly with arsenate anions for available sites, due to the similarities existing between them. This behavior has been reported when studying the co-adsorption of some oxyanions on the goethite surface, positively charged. Paths followed by trivalent oxyanions group (vanadate, phosphate, and arsenate) during adsorption, are practically indistinguishable from each other [27]. In any case, the highest sorption capacity is reached with molybdate-loaded beads without treatment: 1 mmol As g⁻¹ dry mass, which represents an advantage in terms of simplicity and cost of the process.

The conventional models of Langmuir (Eq. 1) and Freundlich (Eq. 2) were used to adjust the experimental data using non-linear regression analysis. Besides, the SIPS model (which combines the Langmuir and the Freundlich

equations, according to Eq. 3) was also tested for modeling the sorption isotherms. The best correlation coefficient is systematically obtained with the SIPS equation, although the values of Langmuir are very close. It is noteworthy that the adjustment of the isotherm profile for tungstate-loaded beads treated with phosphoric acid is very poor, regardless of the fitting model used; this is probably due to the very low sorption capacities obtained with this treated sorbent (i.e., less than 0.19 mmol As g⁻¹).

$$q_{eq} = q_m K_L \frac{c_{eq}}{1 + K_L c_{eq}} \quad (1)$$

$$q_{eq} = K_f C_{eq}^{1/n} \quad (2)$$

$$q_{eq} = \frac{q_m (b c_{eq})^{1/n}}{1 + b c_{eq}^{1/n}} \quad (3)$$

Table 2 shows the parameters determined for each model used. Langmuir equation indicates monolayer sorption on

Table 2 Sorption isotherms—modeling parameters for Langmuir, Freundlich, and SIPS equations

Model	Parameter	Mo	Mo-PO ₄	W	W-PO ₄
Langmuir	$q_{eq,exp}$ (mmol As g ⁻¹ dry mass)	1.007	0.826	0.590	0.0192
	$q_{eq,exp}$ (mmol As g ⁻¹ Mo or W)	116.076	108.122	60.762	21.640
	$q_{eq,cal}$ (mmol As g ⁻¹)	0.927	1.109	0.545	0.194
	K_L (L mmol ⁻¹)	20.743	1.291	15.057	1.901
	R^2	0.964	0.989	0.950	0.800
Freundlich	K_F (mmol g ⁻¹)/(mmol L ⁻¹) ⁿ	0.902	0.593	0.492	0.119
	n	3.507	1.914	3.630	2.405
	R^2	0.939	0.992	0.910	0.803
SIPS	$q_{eq,cal}$ (mmol As g ⁻¹)	1.087	1.908	0.587	0.406
	$1/n$	0.681	0.703	0.804	0.608
	B (mL mmol As ⁻¹)	4.914	0.469	6.867	0.414
	R^2	0.974	0.997	0.952	0.811

the homogeneous surface sites of beads [44]. The equilibrium amount of metal sorbed per unit weight of sorbent is q_e ; the equilibrium concentration of metal in aqueous solution is C_e ; the maximum sorption capacity of the sorbent at saturation of the monolayer is q_m , and K_L is the Langmuir equilibrium coefficient. For composite materials and some chitosan beads, the Freundlich isotherm can also describe the sorption process, where K_F and n are the Freundlich constants, which are indicators of the capacity and intensity of sorption, respectively. However, it can be assumed that chitosan-derived materials present heterogeneities of the sorption sites, and the sorption process could be different from monolayer [45]. The SIPS isotherm model—which gives the best fitting for the equilibrium data— includes three different parameters: q_m is the total number of binding sites, b is the constant average association, and $1/n$ is the factor of heterogeneity. When the factor of heterogeneity is less than one, the sorbent is heterogeneous; but when it is close to the unit, it represents more homogeneous binding sites. This model is more suitable for predicting sorption on heterogeneous surfaces, but if the adsorbate concentration is very low, the model reduces to Freundlich's, and if the adsorbate concentration is high, the model reduces to Langmuir's [46, 47].

Table 3 shows sorption properties on different chitosan-based sorbents. Given the different nature and experimental conditions for each process, it is difficult to make a direct comparison among the sorbents. However, it gives an overview of the potential of molybdate- and tungstate-loaded beads in the removal of As(V). The sorption capacity of CTS-Mo beads is well above several of the compared sorbents; however, there are some reported sorbents with higher

sorption capacity than our material [8, 48–50]. Although this should not be the only parameter to consider: the complexity of materials and manufacturing processes, as well as the necessary sorbent dosage, should also be considered because we are looking for a relatively simple manufacturing process. For example, it is not inexpensive nor straightforward to prepare nanofibers of cellulose acetate/chitosan/SWCNT/Fe₃O₄/TiO₂ [48] or chitosan magnetic material after pyrolysis at 550 °C [49] (pyrolysis will degrade chitosan into char with no NH nor OH functional groups at all). And even the sorption capacity of CTS-W beads is lower than that of CTS-Mo, it is still competitive compared to other chitosan-based sorbents.

Influence of Drying on Sorption Kinetics

Figure 9 compares sorption kinetics for CTS-Mo and CTS-W beads subjected to three different treatments: oven-dried, freeze-dried, and wet (not dried). For both sorbents, the kinetic curve representing the oven-dried process presents a long initial stage where the sorbent is practically unable to uptake metal ions of the solution, which can be explained by the restriction in the porosity of this material. The oven-drying process leads to an irreversible reduction in the size and shape of beads, which in turn causes a decrease in the size and volume of pores that hinder the diffusion process [51]. Even the sorption capacity is not much lower than that of freeze-dried beads, the equilibrium time is considerably longer, needing more than 48 h to reach equilibrium. The kinetic curve for freeze-dried beads present a much faster ion uptake at the beginning, and the equilibrium is reached

Table 3 As(V) sorption capacity comparison for several chitosan-based sorbents

Sorbent	pH range	Sorption capacity (mg g ⁻¹)	Sorbent dosage (mg mL ⁻¹)	Reference
Cellulose acetate/chitosan/SWCNT/Fe ₃ O ₄ /TiO ₂ composite nanofibers	3.0	281.5	0.5	[48]
Chitosan stabilized magnetic material after pyrolysis at 550 °C	6.0	101	4	[49]
Molybdate-loaded chitosan beads	3.0	75	1	This work
Chitosan-functionalized graphene oxide	4.3–6.5	71.9	8	[8]
Cross-linked magnetic chitosan anthranilic acid glutaraldehyde Schiff's base	2.0	62.42	0.5	[50]
Tungstate-loaded chitosan beads	3.0	44	1	This work
Fe–Mn binary oxide impregnated chitosan bead	7.0	39.1	1	[54]
Chitosan/Cu(OH) ₂ composite	4.0	39.1	2.5	[55]
Chitosan/CuO composite	4.0	28.1	2.5	[55]
Chitosan beads immobilized with iron(III)	7.0	27.59	5	[56]
Iron coated chitosan	7.0	22.47 ± 0.56	5	[57]
Granular adsorbent made of iron-containing backwashing residual sludge and chitosan	6.5	14.995	1	[58]
Zeravalent iron impregnated chitosan-cboxymethyl-β-cyclodextrin composite beads	6.0	13.51	3.33	[59]
Chitosan goethite bionanocomposite	5.0–9.0	11.3	2.5	[60]

Fig. 9 Influence of drying process on As(V) sorption. PFORE and PSORE kinetic models for **a** CTS–Mo, and **b** CTS–W beads. Sorbent dosage: 0.8 mg mL^{−1}, pH: 3, C₀: 50 mg As L^{−1}, T: 20 °C

in less than 24 h, favored, in this case, by the high porosity of the sorbent. Despite the favorable effect of freeze-drying on the mass transfer properties, this is an expensive drying process, and the cost–benefit balance should be evaluated for justifying the use of this method. As expected, wet beads show the faster kinetic profiles; this is consistent with conclusions reported on the sorption of dyes on chitosan beads [52]. The first part of the curve has an ion taking almost as fast as freeze-dried beads, and, in the same way, the equilibrium time is less than 24 h; however, its sorption capacity is considerably lower. The above can be attributed mainly to the loss of stability over time since most of the beads get cracked during the process, which also limits their lifetime and prevents reuse.

Pseudo-first (PFORE) and pseudo-second (PSORE) order rate equations (see supplementary material section) were used to adjust the experimental data. These models describe kinetics reactions for homogeneous systems, but they are also commonly used to model sorption kinetics for heterogeneous systems. In general, the pseudo-second-order model predicts the experimental profiles more efficiently. Table 4 shows the parameters determined for each model.

Although PSORE describes adequately kinetic experimental data, the model does not provide information regarding the adsorption mechanism. In such cases, the

intraparticle diffusion model can be useful to identify the mechanism and predict the rate-controlling step. Experimental data is linearized to the intraparticle diffusion model. CTS–Mo and CTS–W beads exhibit two linear regions (Fig. SM4 of the supplementary material), indicating that the sorption process is controlled by a multistep mechanism instead of being entirely governed by intraparticle diffusion [53]. Table SM1(of the supplementary material) shows the parameters calculated for the intraparticle diffusion model.

Effect of Competitor Ions

The influence of other anions (PO_4^{3-} , SO_4^{2-} , NO_3^- , Cl^-) is considered to evaluate the efficiency of the proposed sorbent in the presence of different species from industrial effluents or natural waters. In a second step, the competition of base metals has been investigated using a simulated effluent that contained: Cu(II), Zn(II), Ni(II), and Cd(II). Figure 10 shows the variation in sorption capacity in three different concentrations of co-existing ions. Chloride ions have the least influence on sorption capacity: a considerable excess (i.e., 100:1 ratio) was required for detecting a slight decrease in As(V) binding. Nitrate ions have a limited impact on sorption performance, especially for CTS–W beads: large

Table 4 Sorption kinetics—modeling parameters for PFORE and PSORE kinetic models

Model	Parameter	Mo			W		
		Oven-dried	Freeze-dried	Wet	Oven-dried	Freeze-dried	Wet
PFORE	q_{eq} (mmol As g ^{−1})	0.590	0.654	0.503	0.426	0.428	0.222
	k_f (min ^{−1})	1.38×10^{-3}	8.33×10^{-3}	32.1×10^{-3}	2.00×10^{-3}	20.4×10^{-3}	24.4×10^{-3}
	R ²	0.981	0.976	0.968	0.993	0.972	0.969
PSORE	q_{eq} (mmol As g ^{−1})	0.724	0.701	0.526	0.498	0.448	0.233
	k_2 (g mmol ^{−1} min ^{−1})	1.87×10^{-3}	17.9×10^{-3}	95.9×10^{-3}	4.41×10^{-3}	73.5×10^{-3}	163×10^{-3}
	R ²	0.985	0.995	0.993	0.997	0.989	0.973

Fig. 10 Effect of co-existing anions on As(V) sorption for **a** CTS–Mo and **b** CTS–W beads. Sorbent dosage: 1 mg mL^{−1}, pH: 3, contact time: 140 h, T: 20 °C

excess (i.e., 100:1 ratio) is necessary for a slight decrease in uptake performance. For CTS–Mo beads, the effect of nitrate ions can be detected when the competitor anion is in excess compared to arsenate anions. Sulfates and phosphates are undoubtedly the ions that cause the greatest effect on sorption capacity, this being even greater with phosphate ions. As explained before, phosphate ions share similarities with arsenate ions, and thus compete for sorption sites and reduce its sorption capacity almost completely when the ratio phosphate:arsenate is 100:1. This different effect for other ions present in the solution supports the interpretation of the mechanism as complexation rather than an ion exchange.

The presence of other cations, such as those present in effluents of the mining industry, has a higher impact on arsenic sorption. The two effluents taken as a basis to simulate the actual concentrations of different metals have a high sulfate content respect to arsenic (i.e., 30:1 ratio). Table 5 shows the initial concentration of Cu, Zn, Ni, Cd, and As for both effluents; as can be observed, even when the sorption capacity for other metals is deficient, the arsenic sorption capacity is affected. For CTS–Mo, a maximum

capacity of 0.69 mmol As g^{−1} is reached, while CTS–W beads reach only 0.21 mmol As g^{−1}. This loss of capacity can also be attributed to the high sulfate content, which, as was explained, has the greatest impact on arsenic sorption due to direct competition for active sites.

Arsenic Desorption and Sorbent Reuse

Desorption on CTS–Mo beads had been successfully tested using phosphoric acid 0.1 M [14]; this eluent was also tested for As (V) removal from CTS–W beads. Figure 11 shows the sorption efficiency for beads after three cycles. The CTS–Mo beads show only a slight decrease in sorption capacity in the second cycle, but it is recovered again in the third. The sorption capacity for each cycle is 0.71 ± 0.02 , 0.68 ± 0.01 , and 0.75 ± 0.04 mmol As g^{−1}, respectively. On the opposite side, there is a continuous decrease in the sorption capacity of CTS–W beads throughout the three cycles. The sorption capacity for each cycle is 0.54 ± 0.07 , 0.33 ± 0.01 , and 0.24 ± 0.04 mmol As g^{−1}, respectively. This behavior can be attributed to a progressive saturation of the active sites,

Table 5 Effect of co-existing metal ions on As(V) sorption in a simulated effluent

Effluent	Metal ion	C ₀ (mM)	CTS–Mo beads q _{eq} (mmol As g ^{−1})	CTS–W beads q _{eq} (mmol As g ^{−1})
Effluent 1 (Dziubek [20])	Cu ²⁺	0.202	0.003	0.007
	Zn ²⁺	0.042	0.001	0.004
	Ni ²⁺	0.029	0	0
	As ⁵⁺	3.785	0.694	0.210
Effluent 2 (Basha et al. [19])	Cu ²⁺	0.237	0.008	0.011
	Zn ²⁺	0.318	0.011	0.004
	Ni ²⁺	0.022	0	0.005
	Cd ²⁺	0.051	0.001	0.037
	As ⁵⁺	4.021	0.669	0.211

Fig. 11 Reuse of CTS-Mo and CTS-W chitosan beads. Desorption step—desorption agent: H_3PO_4 0.1 M, sorbent dosage: 0.5 mg mL^{-1} , contact time: 24 h, T: 20°C

considering the desorption efficiency decreases with each cycle. Furthermore, based on the previous treatment with phosphoric acid, there is no evidence that it removes the tungstate bound to chitosan. Tungstate beads were also tested for desorption with 0.1 M NaOH (not shown); after the first and second cycle, the desorption percentage reached is higher than its equivalent with phosphoric acid. However, beads do not remain stable in the solution, after cycles with a considerable sorbent mass loss, which may indicate reversibility in the ionotropic gelation process.

Conclusions

The molybdate and tungstate ions can form complexes with As(V) in solution, which improves the sorption properties of chitosan with arsenic. In this case, and contrary to the previous report for coagulated beads in molybdate solution, additional treatment with phosphoric acid does not improve the sorption capacity of molybdate or tungstate beads. Sorption capacities reached 75 and 44 mg As g^{-1} of dry mass in CTS-Mo and CTS-W beads, respectively. The drying process reveals a higher sorption rate at the beginning of the process for freeze-dried beads; however, when reaching equilibrium, their capacity is not much higher than the capacity reached of oven-dried beads.

Regarding the competition with other ions, the most significant impact on arsenic sorption is caused by the presence of phosphate ions, even at a 1:1 ratio, due to similarities between phosphate and arsenate ions, and thus compete for sorption sites.

Desorption was carried out with phosphoric acid through 3 cycles, CTS-Mo beads presented a better sorption capacity and, therefore, longer reuse time; in contrast to CTS-W beads, which maintained a decrease in sorption capacity throughout the cycles.

CTS-W and CTS-Mo beads are interesting materials for efficient removal of As(V) from acid solutions in the treatment of residual water compared to other chitosan-based sorbents.

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References

- Mohan D, Pittman C Jr (2007) *J Hazard Mater* 142:1–53
- Navarro O, González J, Jénez-Ferreira H, Bautista C-F, Cardona A (2017) *Procedia Eng* 186:333–340
- Missimer TM, Teaf CM, Beeson WT, Maliva RG, Wooschlager J, Covert DJ (2018) *Int J Environ Res Public Health* 15:1–30
- Rahman M, Rahman A, Kaiser Khan M, Renzaho AM (2018) *Ecotoxicol Environ Saf* 150:335–343
- Bhowmick S, Pramanik S, Singh P, Mondal P, Chatterjee D, Nriagu J (2018) *Sci Total Environ* 612:148–169
- Kwok K, Koong LF, Chen G, McKay G (2014) *J Colloid Interface Sci* 416:1–10
- Guibal E (2004) *Sep Purif Technol* 38:43–74
- Kumar AS, Jiang SJ (2016) *J Environ Chem Eng* 4:1698–1713
- Ngah W, Endud C, Mayanar R (2002) *React. Funct. Polym* 50:181–190
- Pang M, Kano N, Imaizumi H (2015) *J Chem Chem Eng* 9:433–441
- Beheshti H, Irani M, Hosseini L, Rahimi A, Aliabadi M (2016) *Chem Eng J* 284:557–564
- Wang Y, Li L, Luo C, Wanga X, Duan H (2016) *Int J Biol Macromol* 86:505–511
- Dambies L, Guibal E, Roze A (2000) *Colloids Surf A* 170:19–31
- Dambies L, Vincent T, Guibal E (2002) *Water Res* 36:3699–3710
- Gibert F, Moine B, Schott J, Dandurand JL (1992) *Contrib Mineral Petrol* 112:371–384
- Rinaudo M (2006) *Prog Polym Sci* 31:603–632
- Kasaai MR (2007) *Carbohydr Polym* 68:477–488
- Brunauer S, Emmett P, Teller E (1938) *J Am Chem Soc* 60:309–319
- Basha CA, Selvi SJ, Ramasamy E, Chellammal S (2008) *Chem Eng J* 141:89–98
- Dziubek J (2017) *Web conf* 17:1–8
- Dambies L, Vincent T, Domard A, Guibal E (2001) *Biomacromol* 2:1454–1463
- Gustafsson JP (2003) *Chem Geol* 200:105–115
- Xu N, Christodoulatos C, Braida W (2006) *Chemosphere* 64:1325–1333
- Koutsospyros A, Braida W, Christodoulatos C, Dermatas D, Strigul N (2006) *J Hazard Mater* 136:1–19
- Rakshit S, Sallman B, Davantès A, Lefèvre G (2017) *Chemosphere* 168:685–691
- Gecol H, Miakatsindila P, Ergican E, Hiibel SR (2006) *Desalination* 197:165–178
- Rietra RP, Hiemstra T, van Riemsdijk WH (1999) *Geochim Cosmochim Acta* 63:3009–3015
- Qian L, Zhang H (2011) *J Chem Technol Biotechnol* 86:172–184
- Rahman I, Vejayakumaran P, Sipaut C, Ismail J, Chee C (2008) *Ceram Int* 2059:2066

30. Di Renzo F, Valentin R, Boissiere M, Tourrette A, Sparapano G, Molvenger K (2005) ... Quignard F. *Chem Mater* 17:4693–4699
31. Robitzer M, Di Renzo F, Quignard F (2011) *Microporous Mesoporous Mater* 140:9–16
32. Klobes P, Meyer K, Munro RG (2006) In: *Specific Surface Area and Pore Analysis by Gas Adsorption* (ed), US Government printing office, Washington DC, p 23–24
33. Instruments Q (2015) *Powder Tech* 51:51
34. Pawlak A, Mucha M (2003) *Thermochim Acta* 396:153–166
35. Fernandes Queiroz M, Teodosio Melo KR, Araujo Sabry D, Lanzi Sasaki G, Oliveira Rocha HA (2015) *Mar Drugs* 13:141–158
36. Tuchowska M, Muir B, Kowalik M, Socha RP, Bajda T (2019) *Materials* 12:2253
37. Guibal E, Milot C, Eterradosi O, Gauffier C, Domard A (1999) *Int J Biol Macromol* 24:49–59
38. Severo E, Rossi Abaide E, Gonçalves Anchieta C, Segabinazzi Foletto V, Trevisan Weber C, Bisognin Garlet T, Foletto E (2016) *Mater Res* 19:781–785
39. Bertoni FA, González JC, García SI, Sala LF, Bellú SE (2018) *Carbohydr Polym* 180:55–62
40. Das J, Sarkar P (2016) *Environ Sci-Wat Res* 2:693–704
41. Alekseev EV, Felbinger O, Wu S, Malcherek T, Depmeier W, Atuchin VV (2013) *J Solid State Chem* 204:59–63
42. Chiban M, Zerbet M, Carja G, Sinan F (2012) *J Environ Chem Ecotoxicol* 4:91–102
43. Taylor JB, Calvert D, Hun IR (1965) *Can J Chem* 43:3045–3051
44. Girma Asere T, Mincke S, Jeriffa DC, Verbeken K, Tessema D, Fufa F (2017) ... Du Laing G. *Int J Environ Res Public Health* 14:1–19
45. Pontoni L, Fabbicino M (2012) *Carbohydr Res* 356:86–92
46. Wan Ngah W, Fatinathan S (2008) *Chem Eng J* 143:62–72
47. Ayawei N, Ebelegi A, Wankasi D (2017) *J Chem* 2017:1–11
48. ZabihiSahebi A, Koushkbaghi S, Pishnamazi M, Askari A, Khosravi R, Irani M (2019) *Int J Biol Macromol* 140:1296–1304
49. Nisticò R, Celi LR, Bianco Prevot A, Carlos L, Magnacca G, Zanzo E, Martin M (2018) *J Hazard Mater* 342:260–269
50. Abou El-Reash YG, Otto M, Kenawy IM, Ouf AM (2011) *Int J Biol Macromol* 49:513–522
51. Ruiz M, Sastre A, Guibal E (2002) *Sep Sci Technol* 37:2143–2166
52. Chiou M, Li H (2003) *Chemosphere* 50:1095–1105
53. Tran HN, You SJ, Hosseini-Bandegharai A, Chao HP (2017) *Water Res* 120:88–116
54. Qi J, Zhang G, Li H (2015) *Bioresour Technol* 193:243–249
55. Elwakeel KZ, Guibal E (2015) *Carbohydr Polym* 134:190–204
56. De Marques Neto JO, Bellato CR, Milagres JL, Pessoa KD, De Alvarenga ES (2013) *J Braz Chem Soc* 24:121–132
57. Gupta A, Chauhan V, Sankaramakrishnan N (2009) *Water Res* 43:3862–3870
58. Zeng H, Yu Y, Wang F, Zhang J, Li D (2020) *Colloids Surf A* 585:124036
59. Tajuddin Sikder M, Tanaka S, Saito T, Kurasaki M (2014) *J Environ Chem Eng* 2:370–376
60. He J, Bardelli F, Gehin A, Silvester E, Charlet L (2016) *Water Res* 101:1–9